

# The impact of elevated CO<sub>2</sub>, increased nitrogen availability and biodiversity on plant tissue quality and decomposition

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## Abstract

Elevated CO<sub>2</sub>, increased nitrogen (N) deposition and increasing species richness can increase net primary productivity (NPP). However, unless there are comparable changes in decomposition, increases in productivity will most likely be unsustainable. Without comparable increases in decomposition nutrients would accumulate in dead organic matter leading to nutrient limitations that could eventually prohibit additional increases in productivity. To address this issue, we measured aboveground plant and litter quality and belowground root quality, as well as decomposition of aboveground litter for one and 2-year periods using *in situ* litterbags in response to a three-way factorial manipulation of CO<sub>2</sub> (ambient vs. 560 ppm), N deposition (ambient vs. the addition of 4 g N m<sup>-2</sup> yr<sup>-1</sup>) and plant species richness (one, four, nine and 16 species) in experimental grassland plots.

Litter chemistry responded to the CO<sub>2</sub>, N and plant diversity treatments, but decomposition was much less responsive. Elevated CO<sub>2</sub> induced decreases in % N and % lignin in plant tissues. N addition led to increases in % N and decreases in % lignin. Increasing plant diversity led to decreases in % N and % lignin and an increase in % cellulose. In contrast to the litter chemistry changes, elevated CO<sub>2</sub> had a much lower impact on decomposition and resulted in only a 2.5% decrease in carbon (C) loss. Detectable responses were not observed either to N addition or to species richness.

These results suggest that global change factors such as biodiversity loss, elevated CO<sub>2</sub> and N deposition lead to significant changes in tissue quality; however, the response of decomposition is modest. Thus, the observed increases in productivity at higher diversity levels and with elevated CO<sub>2</sub> and N fertilization are not matched by an increase in decomposition rates. This lack of coupled responses between production and decomposition is likely to result in an accumulation of nutrients in the litter pool which will dampen the response of NPP to these factors over time.

*Keywords:* biodiversity, decomposition, elevated CO<sub>2</sub>, FACE, litter chemistry, litter quality, nitrogen fertilization, plant chemistry, plant quality

Received 28 June 2006; revised version received 22 December 2006 and accepted 18 April 2007

## Introduction

Carbon (C) and nitrogen (N) content and other aspects of the chemistry of plant tissues (including both the C:N ratio, but also the specific chemistry of C- and N-containing compounds), significantly influence both decomposition and herbivory, two processes that form

the basis for positive and negative feedbacks in ecosystem processes (Meentemeyer, 1978; Melillo *et al.*, 1982). Plant tissue quality (used herein to describe both the stoichiometry and specific chemistry of C- and N-containing compounds), however, can change in response to resource availability, such as N fertilization (Koricheva *et al.*, 1998; Fisk & Fahey, 2001) and elevated CO<sub>2</sub> (Franck *et al.*, 1997; Gahrooe, 1998; Koricheva *et al.*, 1998). In addition, plant tissue quality is sensitive to plant species composition and diversity (Hector *et al.*,

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2000; Knops *et al.*, 2001). Yet, few experiments have examined the interactions among CO<sub>2</sub>, N and plant diversity on ecosystem functioning, despite the increasing evidence that such global change factors can interact with one another (Reich *et al.*, 2001a; Henry *et al.*, 2005).

The impact of resource availability on tissue stoichiometry has been well studied for N additions. Many studies have reported an increase in plant tissue N concentrations as a result of N fertilization (DiTommaso & Aarssen, 1989; Wedin & Tilman, 1996). Elevated CO<sub>2</sub>, in general, decreases plant tissue N concentrations (Koricheva *et al.*, 1998; Lutze *et al.*, 2000; Norby *et al.*, 2001; Cotrufo *et al.*, 2005). Research on the impact of plant species diversity on tissue quality has been rare, but in the few related studies, diversity and N concentrations were found to be negatively correlated (Tilman *et al.*, 1997; Reich *et al.*, 2001a, 2006; Novotny *et al.*, 2007). In contrast to tissue % N, the impact of resource availability on tissue C allocation, or more specifically the relative proportions of soluble carbohydrates, cellulose and lignin in plant tissues, has received much less attention. Elevated CO<sub>2</sub> often leads to an increase in lignin (Koricheva *et al.*, 1998; Penuelas & Estiarte, 1998; Norby *et al.*, 2001), with some exceptions (Cotrufo *et al.*, 2005). N fertilization impacts are not consistent among studies (Ohnmeiss & Baldwin, 1994; Feller, 1996; Hatcher *et al.*, 1997a,b; John & Turkington, 1997). Only a few studies, however, have examined both plant N and C content responses to changes in more than one factor, most often elevated CO<sub>2</sub> and N supply. In these studies, responses appear to be additive and not synergistic (Lutze *et al.*, 2000; Booker & Maier, 2001; King *et al.*, 2001; Henry *et al.*, 2005).

Outside of the elevated CO<sub>2</sub> literature, litter % N has been shown to be a consistent predictor of decomposition rates in many studies (Melillo *et al.*, 1982; Berg & Matzner, 1997), including at the site of the present study (Hobbie, 2005). Hence, decreased tissue N concentrations under elevated CO<sub>2</sub> have been predicted to negatively affect decomposition, but observed decomposition rates under experimentally elevated CO<sub>2</sub> vary greatly, with some studies reporting a positive effect (Franck *et al.*, 1997), some a negative effect (Boerner & Rebbeck, 1995; Scherzer *et al.*, 1998) and some no effect at all (Torbert *et al.*, 1995; Couteaux *et al.*, 1999; Lutze *et al.*, 2000). A meta-analysis by Norby *et al.* (2001) found a pattern of lower tissue N in leaf litter produced under elevated CO<sub>2</sub>, but did not find consistent changes in decomposition rates. Interactions among global change factors on decomposition have received much less attention. Chiefly focusing on increased N addition and elevated CO<sub>2</sub>, these studies have found subtle (Henry *et al.*, 2005) or no interactions (Franck *et al.*, 1997; Lutze *et al.*, 2000; King *et al.*, 2001).

Given both the paucity and mixed nature of the results of studies that have examined global change impacts on plant tissue quality and decomposition, we conducted a long-term field manipulation of global change factors on grassland ecosystem function. Specifically, we determined tissue quality and decomposition in a direct three-way multifactorial manipulation of elevated CO<sub>2</sub> (ambient and 560 ppm), N fertilization (ambient and 4 g N m<sup>-2</sup> yr<sup>-1</sup>) and species diversity (one, four, nine and 16 plant species). We examined four possible ecosystem responses to these manipulations. First, we examined if elevated CO<sub>2</sub>, N fertilization and variation in species diversity led to changes in plant tissue quality both in terms of N content and C allocation. Second, we examined if there were changes in decomposition rates. Third, we examined whether any changes in decomposition rates were caused by tissue quality changes or by changes in the microclimate resulting from changes in plant community composition and structure due to N addition, elevated CO<sub>2</sub> and variation in species diversity. Fourth, we examined if the manipulation of these factors led to additive or complex changes (i.e. significant interactions among the factors) in decomposition rates.

## Methods

This study was part of a free-air CO<sub>2</sub> enrichment (Biocon) experiment at the Cedar Creek Natural History Area in east-central Minnesota, USA. Cedar Creek has a mid-continental climate with hot, humid summers and cold winters. The average date of the last spring freeze is May 9 and the average first fall freeze is September 27. Soils were formed from a sandy glacial outwash and are low in organic matter, N, clay and water-holding capacity (Grigal *et al.*, 1974). The average soil N accumulation was approximately 1.2 g m<sup>-2</sup> yr<sup>-1</sup> from 1983 to 1995, with atmospheric wet N deposition contributing 0.4–0.5 g m<sup>-2</sup> yr<sup>-1</sup> (Knops & Tilman, 2000).

The Biocon study incorporates three treatments: elevated vs. ambient CO<sub>2</sub> (three 20 m diameter rings for each), elevated (4 g N m<sup>-2</sup> yr<sup>-1</sup>) vs. ambient N and four biodiversity treatments planted with one, four, nine or 16 herbaceous plant species. The design consisted of a split plot arrangement of treatments in a completely randomized design with the CO<sub>2</sub> treatment as a whole-plot factor, which was replicated three times among the six rings. The subplot factors of species identity and N treatment were randomly assigned and replicated in individual plots among the six rings (Reich *et al.*, 2001a, 2000). In total there were 371 plots, including 12 bare soil plots. For each of the four combinations of CO<sub>2</sub> and N, there were 32 randomly assigned replicates for the plots planted to one species, 15 for those planted to four

species, 15 for nine species and 12 for 15 species. In addition, 63 plots of four plant species were added to increase the replication to allow testing of functional group diversity (Reich *et al.*, 2004). We used four native or naturalized plant species which commonly occur at Cedar Creek Natural History Area for each functional group. Functional groups and species were C4 grasses *Andropogon gerardii*, *Bouteloua gracilis*, *Schizachyrium scoparium*, *Sorghastrum nutans*; C3 grasses *Agropyron repens*, *Bromus inermis*, *Koeleria cristata*, *Poa pratensis*; forbs *Achillea millefolium*, *Anemone cylindrica*, *Asclepias tuberosa*, *Solidago rigida*; and legumes *Amorpha canescens*, *Lespedeza capitata*, *Lupinus perennis*, *Petalostemum villosum*. CO<sub>2</sub> was applied to maintain an atmospheric concentration of 560 ppm during daylight hours in the growing season (April–October), while N fertilizer was applied three times during the growing season. A total of 371 plots, each 4 m<sup>2</sup>, were planted in 1997, and the elevated CO<sub>2</sub> and N treatments began in April 1998.

#### Tissue and litter collection

Aboveground leaf samples were collected in mid-August 1999 by clipping a 10 cm × 100 cm strip at the soil surface, and sorted into live plant by species and senesced litter (Reich *et al.*, 2001a, 2000). Aboveground tissues were combined by plot and a composite sample for each plot was analyzed for tissue quality. Roots were collected at 0–20 cm depth using three 5 cm wide cores in the same area used for aboveground biomass clipping. Aboveground fully abscised leaf samples were collected from all plots (*N* = 359 plots), except the bare soil plots in August–October of 1999, depending on when each plant species shed its leaves.

#### Litterbags

We used a common substrate of *B. inermis* litter in all plots, including the zero-diversity plots, to assess the impact on decomposition of microclimate differences caused either directly by the treatments, or indirectly by changes in plant composition and biomass, in each plot.

We combined litter from the selected plant species in accordance with their relative abundance for both the litter quality analysis and for the material subsequently placed in the litterbags. We used the August aboveground biomass to determine the relative abundance of each plant species within each plot. It was not feasible to use all plant species because we placed approximately 1 g of plant material in each bag, and we found that the three most abundant plant species comprised approximately 90–100% of the August clipped plant biomass. We constructed three identical sets for each plot, based on the relative abundance of the three most

abundant plant species for each individual plot. One set was used for the initial litter quality, one set was incubated *in situ* for 1 year, and one set was incubated *in situ* for 2 years. In total there were 366 plots, with seven zero-species plots, resulting in 359 plots with litter; however, one bag of the 2001 harvest was damaged by small rodents and excluded from the analysis. These sets of *in situ* litter allowed us to examine the treatment effects on decomposition within each plot.

All litterbags were 5 cm × 10 cm polyester cloth bags (50 µm pore size, Ankom Tech., Fairport, NY, USA) and were filled with approximately 1 g of 2–4-cm-long plant material. Litterbags were placed and staked on the soil surface in the center of each plot starting in November 1999 and collected after 1 year in October 2000, and after 2 years in October 2001.

Because of mineral soil contamination of litterbag samples, we ash corrected all weights. Soils contain on average 0.85% C (Harmon & Lajtha, 1999).

#### Plant quality analysis

All samples were dried to constant dry weight at 55 °C and ground through a 20 mesh with a Wiley mill. C and N were analyzed with a Costech ECS 4010 element analyzer (Costech Analytical Technologies Inc., Valencia, CA, USA). Biomass fractions were determined by sequential extraction following standard forage chemistry methods (Van Soest, 1982). The first extraction (neutral detergent) removes cell contents including soluble carbohydrates, lipids, pectin, starch, soluble protein and nonprotein N. The second fraction (acid detergent) removes hemicellulose and protein bound to cell walls. The third digestion (72% H<sub>2</sub>SO<sub>4</sub>) hydrolyzes and removes cellulose, leaving lignin and related recalcitrant materials as the fourth and final fraction. Although we refer to these fractions by their predominant biochemical components (i.e. soluble carbohydrates, cellulose, hemicellulose and lignin), these fractions are somewhat heterogeneous and not well defined from a strictly biochemical point of view (Van Soest, 1982). Extractions were performed on 0.5 g plant samples placed in acid-resistant heat sealed pouches, which were sequentially extracted in an Ankom fiber analyzer (Ankom Tech.), with samples dried and reweighed after each step.

#### Statistics

All statistics were performed with SPSS 14 for Windows. The CO<sub>2</sub> treatment is nested within ring (1 df), and was tested against the random effect of ring nested within CO<sub>2</sub> (4 df). The main effect of species (15 df), N (1 df) and interactions between CO<sub>2</sub>, N and species diversity

were tested against the residual error (Reich *et al.*, 2001b). Because the CO<sub>2</sub> treatment was tested against the random effect of ring nested within CO<sub>2</sub>, it has a much lower power to detect significant differences. Therefore, we used a *P* value of 0.1 as the critical level to detect significant differences for the CO<sub>2</sub> treatment and a *P* value of 0.05 to detect significant differences in all other treatments and interactions.

The soluble, hemicellulose, cellulose and lignin fractions add up to 100% and a decrease in fraction corresponds with an increase in the other fractions. Consequently, tissue quality measurements are all highly correlated. We used a type III general linear model (GLM) multivariate analysis of variance to examine the overall impact of species diversity, CO<sub>2</sub> and N treatment on tissue quality. Subsequently, we used a type III univariate GLM to determine which quality factor caused the overall significance. We also used the GLM procedure to estimate the marginal means for each factor (i.e. species, CO<sub>2</sub> and N levels).

We only present C mass loss throughout the paper because mass loss showed the same identical patterns as C loss. Initial % C and % N of the common substrate, *B. inermis*, was determined from 10 replicate samples. The initial % C and % N of each litter sample was determined on a composite sample for each individual plot (see 'Litterbags'). Percent C loss was calculated as  $((\text{initial \% C} \times \text{initial litter sample weight}) - (\text{final \% C} \times \text{final litter sample weight})) / (\text{initial \% C} \times \text{initial litter sample weight}) \times 100$ . Percent N loss was calculated similarly.

Lastly, we used plant tissue N concentration throughout the manuscript for N quality. We also calculated C:N ratio of plant and litter tissues, but the C:N ratio

showed the exact same significance of % N, except its correlation with other factors, or *P* value, was slightly lower.

## Results

### Tissue quality

All three treatments (i.e. CO<sub>2</sub>, N and species diversity) significantly affected aboveground biomass, belowground biomass and litter quality. However, there was no interaction among any of the treatments (Table 1). Some tissue quality metrics were highly correlated: % N was negatively correlated with % hemicellulose and cellulose, and positively correlated with % soluble (except in the root tissues) and % lignin (Table 2).

Elevated CO<sub>2</sub> decreased % N in aboveground biomass and litter (Table 3, Fig. 1a). Elevated CO<sub>2</sub> also decreased the % lignin in the roots and litter (Table 3, Fig. 1a), increased the % soluble (control 30.9%, SE 0.9%, elevated CO<sub>2</sub> 33.9%, SE 0.9%) and decreased the % hemicellulose (control 28.2%, SE 0.6%, elevated CO<sub>2</sub> 26.9%, SE 0.6%) in the belowground biomass (Table 3). The N treatment increased the % N in all three tissue types and decreased the % lignin in the aboveground biomass (Table 3, Fig. 1b). Increasing species diversity treatment decreased the % N and lignin in all three tissues, and increased the % cellulose in both the aboveground biomass and the litter (Table 3, Fig. 1c).

### Decomposition

We found a significant effect of diversity on *B. inermis*, the common substrate litter, when all plots were

**Table 1** Tissue quality: multivariate analysis of aboveground biomass, root and litter with as quality aspects % N, % soluble, % hemicellulose, % cellulose and % lignin and as independent factors CO<sub>2</sub> treatment, N treatment and plant diversity

Treatment (df)	Biomass		Roots		Litter	
	<i>F</i>	<i>P</i>	<i>F</i>	<i>P</i>	<i>F</i>	<i>P</i>
CO <sub>2</sub> (1, 4)	1.6	0.040	1.6	0.058	2.1	0.004
N (1, 338)	4.8	0.000	4.7	0.001	3.1	0.010
Species diversity (3, 338)	9.4	0.000	3.1	0.000	10.0	0.000
CO <sub>2</sub> × N (1, 338)	0.7	0.621	0.5	0.701	0.3	0.922
CO <sub>2</sub> × species (3, 338)	0.6	0.850	0.6	0.835	0.4	0.978
N × species (3, 338)	0.9	0.531	0.8	0.610	0.8	0.641
CO <sub>2</sub> × N × species (3, 338)	0.6	0.901	0.6	0.879	0.4	0.989

The CO<sub>2</sub> is nested within ring. Given are the *F* value of the Pillai's trace and the *P* value. (Pillai's trace and Roy's largest root gave the same results, except for the root CO<sub>2</sub> where the Roy's largest root gave an *F* value of 4.4 and a *P* value of 0.002.) Pillai's trace is more robust to violations of assumptions, whereas Roy's largest root has the greatest power (Scheiner, 2001). All the litter data, the % N, the soluble and lignin for aboveground biomass and the % N for roots are Ln transformed to improve normality.

**Table 2** Correlations among tissue quality measures in aboveground biomass (A), belowground biomass (B) and (C) litter

	% C	% N	% soluble	% hemicellulose	% cellulose
<i>(A) Aboveground biomass</i>					
% N	0.069 <sup>ns</sup>				
% soluble	-0.258 <sup>***</sup>	0.277 <sup>***</sup>			
% hemicellulose	0.074 <sup>ns</sup>	-0.333 <sup>***</sup>	-0.698 <sup>***</sup>		
% cellulose	0.343 <sup>***</sup>	-0.336 <sup>***</sup>	-0.800 <sup>***</sup>	0.379 <sup>***</sup>	
% lignin	0.181 <sup>**</sup>	0.413 <sup>***</sup>	0.268 <sup>***</sup>	-0.770 <sup>***</sup>	-0.116*
<i>(B) Belowground biomass</i>					
% N	0.209 <sup>***</sup>				
% soluble	-0.425 <sup>***</sup>	0.064 <sup>ns</sup>			
% hemicellulose	0.356 <sup>***</sup>	-0.129*	-0.914 <sup>***</sup>		
% cellulose	0.325 <sup>***</sup>	-0.034 <sup>ns</sup>	-0.807 <sup>***</sup>	0.596 <sup>***</sup>	
% lignin	0.393 <sup>***</sup>	0.176 <sup>**</sup>	-0.514 <sup>***</sup>	0.288 <sup>***</sup>	0.315 <sup>***</sup>
<i>(C) Litter</i>					
% N	0.061 <sup>ns</sup>				
% soluble	-0.327 <sup>***</sup>	0.519 <sup>***</sup>			
% hemicellulose	0.211 <sup>***</sup>	-0.505 <sup>***</sup>	-0.880 <sup>***</sup>		
% cellulose	0.239 <sup>***</sup>	-0.656 <sup>***</sup>	-0.854 <sup>***</sup>	0.708 <sup>***</sup>	
% lignin	0.100 <sup>ns</sup>	0.558 <sup>***</sup>	0.496 <sup>***</sup>	-0.724 <sup>***</sup>	-0.497 <sup>***</sup>

Sample size is 359 for each analysis.

<sup>ns</sup> $P > 0.05$ , \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.0001$ .

The C:N ratio, which would show the opposite of the % N, is not shown.

C, carbon; N, nitrogen.

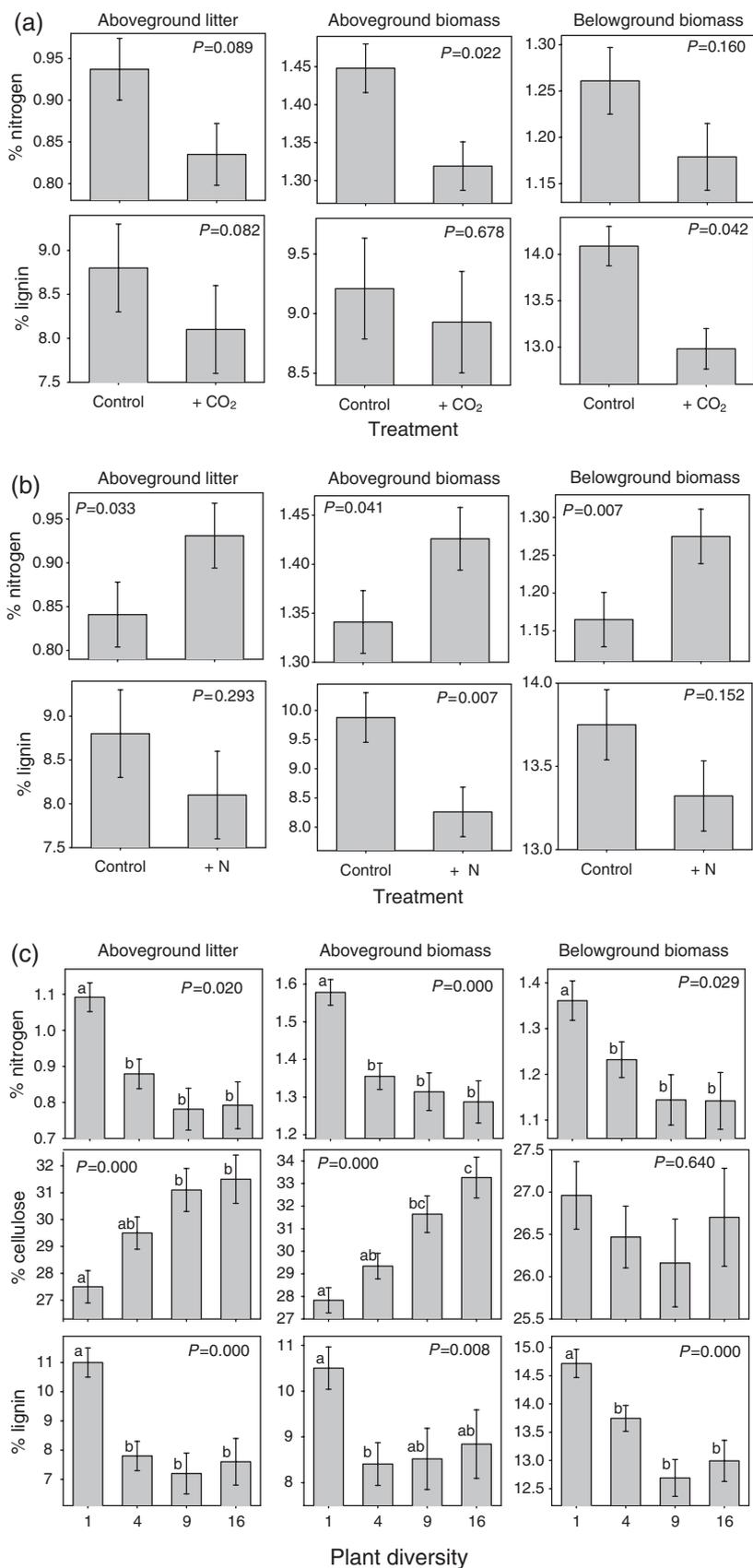
**Table 3** Tissue quality: ANOVAs of aboveground biomass (A), belowground biomass (B) and litter (C) with as quality aspects % N, % soluble, % hemicellulose, % cellulose and % lignin and as independent factors CO<sub>2</sub> treatment, N treatment and plant diversity

Treatment (df)	% N		% Soluble		% hemicellulose		% cellulose		% lignin	
	F	P	F	P	F	P	F	P	F	P
<i>(A) Aboveground biomass</i>										
CO <sub>2</sub> (1, 4)	13.2	0.022	0.1	0.721	0.9	0.396	0.2	0.647	0.2	0.678
N (1, 347)	4.2	0.041	0.2	0.682	2.0	0.161	0.2	0.661	7.3	0.007
Species diversity (4, 347)	6.9	0.000	0.8	0.513	0.8	0.511	11.0	0.000	4.0	0.008
<i>(B) Belowground biomass</i>										
CO <sub>2</sub> (1, 4)	3.0	0.160	7.3	0.054	6.2	0.068	1.0	0.384	8.6	0.042
N (1, 347)	7.5	0.007	0.5	0.480	0.2	0.669	2.9	0.090	2.1	0.152
Species diversity (4, 347)	3.0	0.029	1.5	0.210	0.7	0.392	0.6	0.640	9.9	0.000
<i>(C) Litter</i>										
CO <sub>2</sub> (1, 4)	5.0	0.089	0.0	0.873	0.0	0.889	1.8	0.256	5.3	0.082
N (1, 347)	4.6	0.033	0.0	0.931	0.7	0.387	0.2	0.679	1.1	0.293
Species diversity (4, 347)	3.3	0.020	0.7	0.574	2.1	0.105	6.6	0.000	10.0	0.000

Given are the *F* and *P* values from a GLM with as factors CO<sub>2</sub> nested within ring, N and plant species diversity. Percent N was Ln transformed to improve normality.

N, nitrogen; GLM, general linear model.

**Fig. 1** Tissues quality responses to manipulated global change factors. (a) Responses to elevated CO<sub>2</sub>. (b) Responses to increased nitrogen (N) deposition. (c) Responses to manipulated levels of plant species richness. Note that there were no significant interactions among treatments (Table 1). Shown are the means adjusted for the other treatments  $\pm$  standard error for each treatment effects on % N and lignin for the CO<sub>2</sub> and N treatments and % N, cellulose and lignin for the diversity treatment. Different letters denote  $P < 0.05$ , Bonferroni corrected for multiple comparisons (see Table 3).



**Table 4** Decomposition: ANOVAs of carbon (C) and nitrogen (N) loss over a 1- and a 2-year period, with as independent factors CO<sub>2</sub> treatment, N treatment and plant diversity of (A) a common substrate (*Bromus inermis*) litter, (B) *Bromus* litter, with the bare ground plots excluded and (C) the *in situ* plot representative litter

Treatment (df)	C loss				N loss			
	2000		2001		2000		2001	
	F	P	F	P	F	P	F	P
<i>(A) Bromus litter</i>								
CO <sub>2</sub> (1, 4)	0.0	0.854	2.2	0.216	0.2	0.647	0.3	0.598
N (1, 347)	2.2	0.140	0.8	0.367	1.7	0.193	2.0	0.157
Species diversity (4, 347)	4.2	0.002	7.8	0.000	3.8	0.005	3.5	0.008
CO <sub>2</sub> × N (1, 347)	0.0	0.837	0.1	0.761	0.2	0.699	0.5	0.467
CO <sub>2</sub> × species (4, 347)	0.5	0.721	0.7	0.582	0.2	0.944	2.1	0.079
N × Species (4, 347)	1.2	0.314	0.9	0.481	0.3	0.863	0.5	0.724
CO <sub>2</sub> × N × species (4, 347)	0.6	0.649	2.6	0.036	0.4	0.795	0.9	0.476
<i>(B) Bromus litter, no bare ground plots</i>								
CO <sub>2</sub> (1, 4)	0.0	0.963	1.9	0.241	0.6	0.496	0.1	0.723
N (1, 339)	0.0	0.927	1.8	0.184	2.1	0.145	1.6	0.205
Species diversity (3, 339)	0.3	0.845	1.4	0.229	1.5	0.217	2.4	0.068
CO <sub>2</sub> × N (1, 339)	1.1	0.303	3.6	0.059	0.2	0.667	4.3	0.040
CO <sub>2</sub> × species (3, 339)	0.6	0.586	0.6	0.585	0.2	0.871	2.1	0.096
N × species (3, 339)	0.5	0.716	1.2	0.326	0.4	0.731	0.6	0.606
CO <sub>2</sub> × N × species (3, 339)	0.7	0.572	2.9	0.035	0.6	0.624	0.8	0.497
<i>(C) In situ, plot representative litter</i>								
CO <sub>2</sub> (1, 4)	3.6	0.129	7.2	0.055	0.0	0.839	0.2	0.652
N (1, 339)	2.7	0.104	0.6	0.452	0.0	0.949	0.4	0.538
Species diversity (3, 339)	13.9	0.000	11.4	0.000	0.5	0.661	1.0	0.387
CO <sub>2</sub> × N (1, 339)	0.0	0.957	0.1	0.793	0.4	0.507	0.0	0.829
CO <sub>2</sub> × species (3, 339)	2.3	0.081	0.7	0.535	0.4	0.744	1.1	0.332
N × species (3, 339)	0.2	0.878	0.0	0.987	0.2	0.925	0.1	0.978
CO <sub>2</sub> × N × species (3, 339)	0.3	0.830	0.2	0.895	0.3	0.835	0.1	0.951

Given are the *F* values from a GLM with as factors CO<sub>2</sub> nested within ring, N and plant species diversity. All data Ln transformed. GLM, general linear model.

included (Table 4a, Fig. 2), but no diversity effect when the zero-diversity plots were excluded from the analysis (Table 4b). Excluding the zero-diversity plots, in year 1 we found that there were no significant main effects of CO<sub>2</sub> or N on decomposition, nor a significant interaction (Table 4b). The N loss in the second year showed a significant CO<sub>2</sub> × N interaction (Table 4b) caused by a 3% lower N loss from the elevated CO<sub>2</sub> and N fertilized treatment; but this was not matched by a similar difference in C release (Fig. 3). Within the monocultures, there were also no significant differences among the functional groups (all *P* > 0.2).

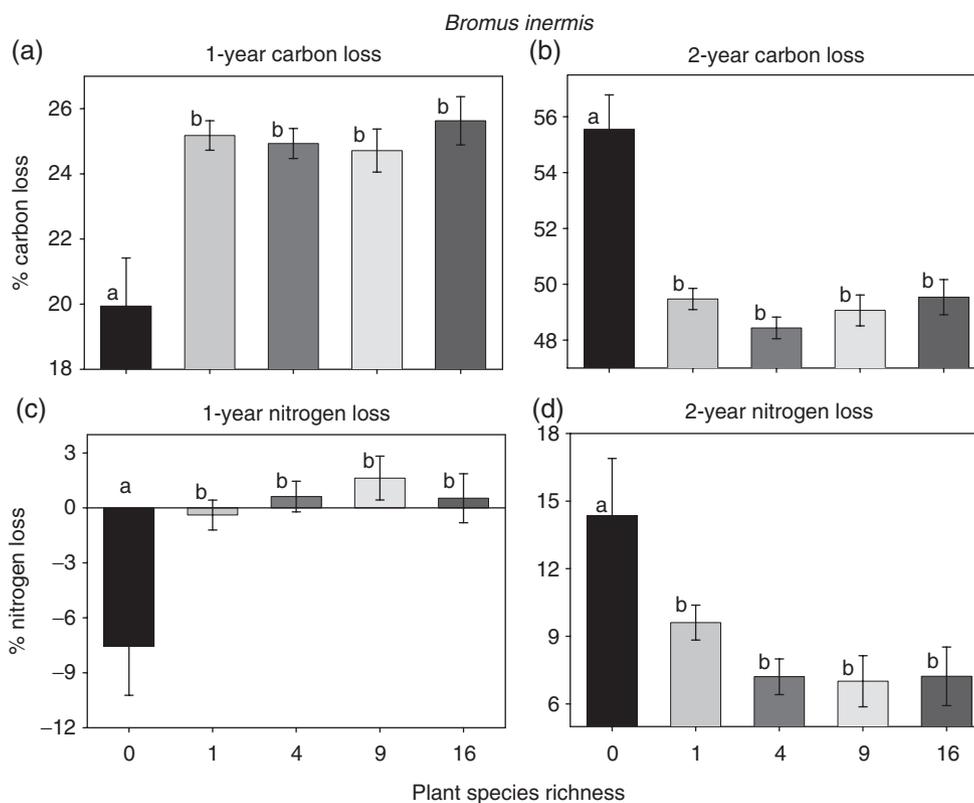
Litter, representative of the plot and decomposed *in situ*, showed no significant change in N loss. The 1-year litter immobilized on average 20.6% of its original N amount and the 2-year litter immobilized on average 8.9% of its original N content. C did show an absolute 2.5% lower C loss rate under elevated CO<sub>2</sub> in

both years (Fig. 4), but this was only significant over the 2 years (Table 4c). However, this decomposition change was relatively small compared with the overall C loss [i.e. 33.0% in the control and 31.1% in the elevated CO<sub>2</sub> treatment in the first year, and 54.8% in the control and 52.3% in the elevated CO<sub>2</sub> treatment over 2 years (Fig. 4)]. There was a significant increase in C loss rate with increasing species richness (Table 4c), which was driven by the monoculture plots, which on average lost about 5% less C as compared with all other diversity levels (Fig. 5).

## Discussion

### Tissue quality

We found significant and similar correlations between the components of tissue quality for above- and below-



**Fig. 2** Percent carbon (C) loss (a, b) and % nitrogen (N) loss (c, d) of a common substrate litter (*Bromus inermis*) over 1- and 2-year periods vs. plant species richness. Shown are the means for each species richness treatment, adjusted for the CO<sub>2</sub> and N treatments  $\pm$  standard error. Different letters denote  $P > 0.05$  (see Table 4a).

ground biomass and litter, consistent with earlier studies (Norby *et al.*, 2000; Hoorens *et al.*, 2003). This consistency indicates that resource induced changes in plant tissue quality are also reflected in changes in litter quality, and that changes in green and senesced tissue chemistry changes correspondingly.

Within this study, we found no significant interactions among elevated CO<sub>2</sub>, N fertilization and changes in plant species diversity on plant tissue quality. Thus, the impact of elevated CO<sub>2</sub>, N fertilization and biodiversity on tissue quality are independent of one another within this study. Tissue N concentrations increased with N fertilization and decreased with elevated CO<sub>2</sub>, but the latter was only significant for the belowground biomass. The observed shifts in tissue C chemistry due to elevated CO<sub>2</sub> and N fertilization were also consistent with prior studies (Ohnmeiss & Baldwin, 1994; Feller, 1996; Hatcher *et al.*, 1997a, b; John & Turkington, 1997; Penuelas & Estiarte, 1998; Norby *et al.*, 2001).

The species richness treatments had the strongest influence on the litter quality ( $F$  value of 10.0 vs. 2.1 for CO<sub>2</sub> and 3.1 for N), and the litter quality showed a significant decrease in % N and lignin and an increase in cellulose with increasing plant diversity.

### Decomposition

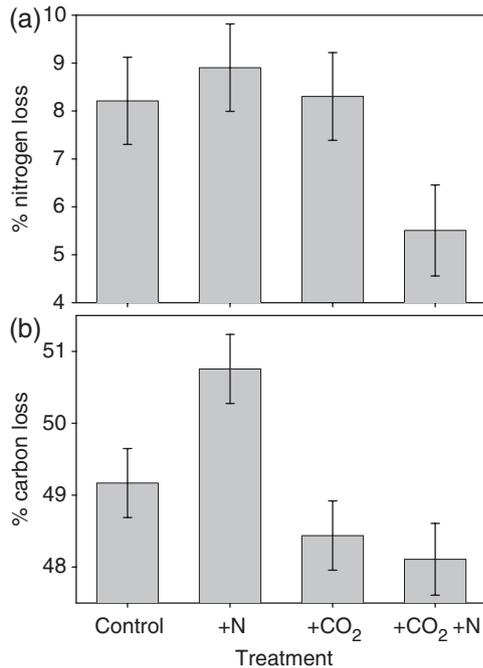
Elevated CO<sub>2</sub>, N fertilization and different levels of diversity can impact decomposition in two ways. First, resource changes can lead to changes in the vegetation structure and the standing biomass (Reich *et al.*, 2001a), which in turn change the microclimate and communities of soil microbial decomposers communities (Epstein *et al.*, 2002; Waldrop & Firestone, 2004). Second, resource changes can lead to changes in tissue quality.

Our common substrate showed no effect on its decomposition rate from either N fertilization, elevated CO<sub>2</sub> or any interactions among the treatments. The lack of a N fertilization effect is similar to other studies with a similar range of N additions (Knorr *et al.*, 2005). The lack of a CO<sub>2</sub> response for decomposition of the common substrate contrasts with the CO<sub>2</sub> response of productivity which resulted in changes in aboveground biomass (Reich *et al.*, 2001a). Decomposition rates were lower in bare soil plots and elevated in the second year, but we found no significant differences at any diversity level at which plants were present. This lack of a diversity effect is consistent with results from a common litter within a similar biodiversity experiment at

the same site of plots that differed twofold in biomass (Knops *et al.*, 2001). Thus, there is no indication that changes in the vegetation structure and the standing biomass (Reich *et al.*, 2001a) have any impact on decomposition.

The litter produced within a plot, which did show changes in N concentration and C fractions, also showed changes in decomposition, but these changes were relatively minor and smaller than litter and plant biomass changes (Reich *et al.*, 2001a). Corresponding

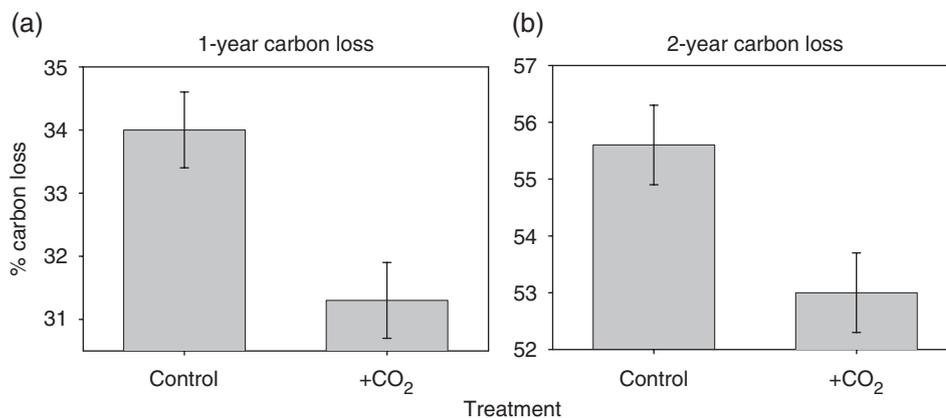
with the plant and litter tissue quality, we found no interactions among our treatments on decomposition. The C loss was 2.5% lower under elevated CO<sub>2</sub>, both after 1 and 2 years, and increased 5% from the monoculture to the other diversity levels. Surprisingly, we found no significant effect of the N treatment, even though the N treatment led to higher tissue N concentrations. Thus, even though there are consistent differences in litter quality, litter quality only has a minor influence on C loss rates and no impact on N loss rates. Likewise, other studies have found that elevated CO<sub>2</sub> (Norby *et al.*, 2001; Hoorens *et al.*, 2003) and N fertilization (Knorr *et al.*, 2005) can lead to plant tissue changes, but often do not lead to changes in decomposition.



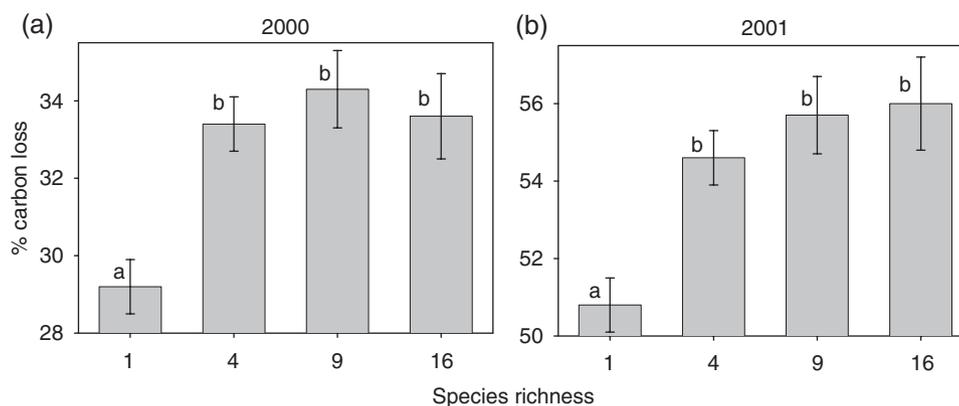
**Fig. 3** Percent nitrogen (N) loss (a) and % carbon (C) loss (b) of the common substrate litter (*Bromus inermis*) over a 2-year period. Shown are the means for each CO<sub>2</sub> × N treatment adjusted for the species richness treatments ± standard error (see Table 4b).

#### *Ecosystem feedbacks through tissue quality changes and decomposition*

This study supports the view (Finzi & Schlesinger, 2002; Weatherly *et al.*, 2003; Allard *et al.*, 2004; Henry *et al.*, 2005) that species compositional changes are much more important in determining feedbacks through N cycling, as compared with any direct or indirect change of species tissue quality change caused by either CO<sub>2</sub> or N fertilization. We found only minor changes of litter decomposition and no interactions between N fertilization and elevated CO<sub>2</sub>. These and similar results (Cotrufo *et al.*, 1994; Gahrooe, 1998; Norby *et al.*, 2001; Finzi & Schlesinger, 2002; Hoorens *et al.*, 2003; de Graaff *et al.*, 2004) are increasing indications that there are none or only minor qualitative changes in litter decomposition due to elevated CO<sub>2</sub> (Zak *et al.*, 2003; de Graaff *et al.*, 2006). Therefore, feedbacks of tissue quality due to elevated CO<sub>2</sub> on N cycling seem unlikely. This study also shows that N fertilization under elevated CO<sub>2</sub> does not necessarily lead to increased decomposition rates, as we found no interactions for tissue quality



**Fig. 4** *In situ* % carbon (C) loss of a plot representative aboveground litter sample over a 1- (a) and 2-year (b) period. Shown are the means for each CO<sub>2</sub> treatment, adjusted for the nitrogen (N) and species richness treatments ± standard error (see Table 4c).



**Fig. 5** *In situ* % carbon (C) loss of a plot representative aboveground litter sample over a 1- (a) and 2-year (b) period. Shown are the means for the each species richness treatment, adjusted for the CO<sub>2</sub> and nitrogen (N) treatments  $\pm$  standard error. Different letters denote  $P > 0.05$  (see Table 4c).

or for decomposition, and N fertilization did not result in increased decomposition rates.

Productivity and biomass accumulation within this experiment, however, did increase in response to CO<sub>2</sub> and N as has been shown in the majority of elevated CO<sub>2</sub> studies (Luo *et al.*, 2006) and showed a larger response in species rich plots (Reich *et al.*, 2001a). Thus, there is an increase in litter inputs both with single global change factor responses and with a significant additive interaction among these factors. Quantitatively, this can lead to a negative feedback because the increased litter pool immobilizes 21% of its initial N content over 1 year and 9% over 2 years of decomposition, as this study shows. This may, over time with annual additions of different litter amounts, lead to a decrease in N availability and a negative feedback on productivity if a global change factor leads to increased productivity. Such a feedback has the potential to diminish any direct impact of global change on ecosystem productivity. Recent productivity data from this experiment shows that the initial increased productivity with elevated CO<sub>2</sub> has decreased (Reich *et al.*, 2006).

### Acknowledgements

This research was supported by the US Department of Energy and the National Science Foundation (NSF) Long-Term Ecological Research (DEB-0080382) and Biocomplexity (DEB-0322057) programs. We thank Jared Trost for help in the field, Chelsea Philippe for tissue analysis, Cathleen Mcfadden for C and N analysis, Amy Kochsiek, Katie Potter and Dave Wedin for comments.

### References

Allard V, Newton PCD, Liefvering M, Soussana JF, Grieu P, Matthews C (2004) Elevated CO<sub>2</sub> effects on decomposition

processes in a grazed grassland. *Global Change Biology*, **10**, 1553–1564.

Berg B, Matzner E (1997) Effect of N deposition of plant litter and soil organic matter in forest systems. *Environmental Reviews*, **5**, 1–25.

Boerner REJ, Rebeck J (1995) Decomposition and nitrogen release from leaves of three hardwood species grown under elevated O<sub>3</sub> and/or CO<sub>2</sub>. *Plant and Soil*, **170**, 149–157.

Booker FL, Maier CA (2001) Atmospheric carbon dioxide, irrigation, and fertilization effects on phenolic and nitrogen concentrations in loblolly pine (*Pinus taeda*) needles. *Tree Physiology*, **21**, 609–616.

Cotrufo MF, de Angelis P, Polle A (2005) Leaf litter production and decomposition in a poplar short-rotation coppice exposed to free air CO<sub>2</sub> enrichment (POPFACE). *Global Change Biology*, **11**, 971–982.

Cotrufo MF, Ineson P, Rowland AP (1994) Decomposition of tree leaf litter grown under elevated CO<sub>2</sub>: effect of litter quality. *Plant and Soil*, **163**, 121–130.

Couteaux MM, Kurz C, Bottner P, Raschi A (1999) Influence of increased atmospheric CO<sub>2</sub> concentration on quality of plant material and litter decomposition. *Tree Physiology*, **19**, 301–311.

de Graaff MA, Six J, Blum H, van Kessel C (2006) Prolonged elevated atmospheric CO<sub>2</sub> does not affect decomposition of plant material. *Soil Biology and Biochemistry*, **38**, 187–190.

de Graaff MA, Six J, Harris D, Blums H, Van Kessel C (2004) Decomposition of soil and plant carbon from pasture systems after 9 years of exposure to elevated CO<sub>2</sub>: impact on C cycling and modeling. *Global Change Biology*, **10**, 1922–1935.

DiTommaso A, Aarssen LW (1989) Resource manipulations in natural vegetations: a review. *Vegetation*, **84**, 9–29.

Epstein HE, Parker IC, Lauenroth WK (2002) Regional patterns of decomposition and primary production rates in the US Great Plains. *Ecology*, **83**, 320–327.

Feller IC (1996) Effects of nutrient enrichment on growth and herbivory of dwarf red mangrove (*Rhizophora mangle*). *Ecological Monographs*, **65**, 477–505.

Finzi AC, Schlesinger WH (2002) Species control variation in litter decomposition in a pine forest exposed to elevated CO<sub>2</sub>. *Global Change Biology*, **8**, 1217–1229.

- Fisk MC, Fahey TJ (2001) Microbial biomass and nitrogen cycling responses to fertilization and litter removal in young northern hardwood forests. *Biogeochemistry*, **53**, 201–223.
- Franck VM, Hungate BA, Chapin FS III, Field CB (1997) Decomposition of litter produced under elevated CO<sub>2</sub>: dependence on plant species and nutrient supply. *Biogeochemistry*, **36**, 223–237.
- Gahrooe FR (1998) Impacts of elevated CO<sub>2</sub> on litter quality, litter decomposability and nitrogen turnover rate of two oak species in a Mediterranean forest ecosystem. *Global Change Biology*, **4**, 667–677.
- Grigal DF, Chamberlain LM, Finney HR, Wroblewski DW, Gross ER (1974) *Soils of the Cedar Creek Natural History Area*. University of Minnesota Agriculture Experiment Station, St Paul.
- Harmon ME, Lajtha K (1999) Analysis of detritus and organic horizons for mineral and organic constituents. In: *Standard Soil Methods for Long-Term Ecological Research* (eds Robertson GP, Coleman DC, Bledsoe CS, Sollins P), pp. 143–165. Oxford University Press, New York.
- Hatcher PE, Paul ND, Ayres PG, Whittaker JB (1997a) The effect of nitrogen fertilization and rust fungus infection, singly and combined, on the leaf chemical composition of *Rumex obtusifolius*. *Functional Ecology*, **11**, 545–553.
- Hatcher PE, Paul ND, Ayres PG, Whittaker JB (1997b) Nitrogen fertilization affects interactions between the components of an insect–fungus–plant tripartite system. *Functional Ecology*, **11**, 537–544.
- Hector A, Beale AJ, Minns A, Otway SJ, Lawton JH (2000) Consequences of the reduction of plant diversity for litter decomposition: effects through litter quality and microenvironment. *Oikos*, **90**, 357–371.
- Henry HAL, Cleland EE, Field CB, Vitousek PM (2005) Interactive effects of elevated CO<sub>2</sub>, N deposition and climate change on plant litter quality in a California annual grassland. *Oecologia*, **142**, 465–473.
- Hobbie SE (2005) Contrasting effects of substrate and fertilizer nitrogen on the early stages of litter decomposition. *Ecosystems*, **8**, 644–656.
- Hoorens B, Aerts R, Stroetenga M (2003) Is there a trade-off between the plant's growth response to elevated CO<sub>2</sub> and subsequent litter decomposability? *Oikos*, **103**, 17–30.
- John E, Turkington R (1997) A 5-year study of the effects of nutrient availability and herbivory on two boreal forest herbs. *Journal of Ecology*, **85**, 419–430.
- King JS, Pregitzer KS, Zak DR, Kubiske ME, Holmes WE (2001) Correlation of foliage and litter chemistry of sugar maple, *Acer saccharum*, as affected by elevated CO<sub>2</sub> and varying N availability, and effects on decomposition. *Oikos*, **94**, 403–416.
- Knops JMH, Tilman D (2000) Dynamics of soil carbon and nitrogen accumulation for 61 years after agricultural abandonment. *Ecology*, **81**, 88–98.
- Knops JMH, Wedin D, Tilman D (2001) Biodiversity and decomposition in experimental grassland ecosystems. *Oecologia*, **126**, 429–433.
- Knorr M, Frey SD, Curtis OS (2005) Nitrogen additions and litter decomposition: a meta-analysis. *Ecology*, **86**, 3252–3257.
- Koricheva J, Larsson S, Haukioja E, Keinanen M (1998) Regulation of woody plant secondary metabolism by resource availability: hypothesis testing by means of meta-analysis. *Oikos*, **83**, 212–226.
- Luo Y, Hui D, Zhang D (2006) Elevated CO<sub>2</sub> stimulates net accumulations of carbon and nitrogen in land ecosystems: a meta-analysis. *Ecology*, **87**, 53–63.
- Lutze JL, Gifford RM, Adams HN (2000) Litter quality and decomposition in *Danthonia richardsonii* swards in response to CO<sub>2</sub> and nitrogen supply over four years of growth. *Global Change Biology*, **6**, 13–24.
- Meentemeyer V (1978) Macroclimate and lignin control of litter decomposition rates. *Ecology*, **59**, 465–472.
- Melillo JM, Aber JD, Muratore JF (1982) Nitrogen and lignin control of hardwood leaf litter decomposition dynamics. *Ecology*, **63**, 621–626.
- Norby RJ, Cotrufo MF, Ineson P, O'Neill EG, Canadell JG (2001) Elevated CO<sub>2</sub>, litter chemistry, and decomposition: a synthesis. *Oecologia*, **127**, 153–165.
- Norby RJ, Long TM, Hartz-Rubin JS, O'Neill EG (2000) Nitrogen resorption in senescing tree leaves in a warmer CO<sub>2</sub> enriched atmosphere. *Plant and Soil*, **224**, 15–29.
- Novotny AM, Schade JD, Hobbie SE, Kay AD, Kyle M, Reich PB, Elser JJ (2007) Stoichiometric response of nitrogen-fixing and non-fixing dicots to manipulations of CO<sub>2</sub>, nitrogen, and diversity. *Oecologia*, **151**, 687–696.
- Ohmmeiss TE, Baldwin IT (1994) The allometry of nitrogen allocation to growth and an inducible defense under nitrogen-limited growth. *Ecology*, **75**, 995–1002.
- Penuelas J, Estiarte M (1998) Can elevated CO<sub>2</sub> affect secondary metabolism and ecosystem function? *Trends in Ecology and Evolution*, **13**, 20–24.
- Reich P, Knops J, Tilman D *et al.* (2001a) Plant diversity enhances ecosystem responses to elevated CO<sub>2</sub> and nitrogen deposition. *Nature*, **410**, 809–812.
- Reich P, Tilman D, Craine J *et al.* (2001b) Do species and functional groups differ in acquisition and use of C, N, and water under varying atmospheric CO<sub>2</sub> and N deposition regimes? A field test with 16 grassland species. *New Phytologist*, **150**, 435–448.
- Reich PB, Hobbie SE, Lee T *et al.* (2006) Nitrogen limitation constrains sustainability of ecosystem response to CO<sub>2</sub>. *Nature*, **440**, 922–925.
- Reich PB, Tilman D, Naeem S, Ellsworth DS, Knops J, Craine J, Wedin D, Trost J (2004) Species and functional group diversity independently influence biomass accumulation and its response to CO<sub>2</sub> and N. *Proceedings of the National Academy of Sciences of the United States of America*, **101**, 10101–10106.
- Scheiner SM (2001) MANOVA: multiple response variables and multispecies interactions. In: *Design and Analysis of Ecological Experiments* (eds Scheiner SM, Gurevitch J), pp. 99–115. Oxford University Press, Oxford.
- Scherzer AJ, Rebeck J, Boerner REJ (1998) Foliar nitrogen dynamics and decomposition of yellow-poplar and eastern white pine during four seasons of exposure to elevated ozone and carbon dioxide. *Forest Ecology and Management*, **109**, 355–366.
- Tilman D, Knops J, Wedin D, Reich P, Ritchie M, Siemann E (1997) The influence of functional diversity and composition on ecosystem processes. *Science*, **277**, 1300–1302.

- Torbert HA, Prior SA, Rogers HH (1995) Elevated atmospheric carbon dioxide effects on cotton plant residue decomposition. *Soil Science Society of America Journal*, **59**, 1321–1328.
- Van Soest PJ (1982) *Nutritional Ecology of the Ruminant*. Cornell University Press, Ithaca, NY.
- Waldrop MP, Firestone MK (2004) Microbial community utilization of recalcitrant and simple carbon compounds: impact of oak-woodland plant communities. *Ecologia*, **138**, 275–284.
- Weatherly HE, Zitzer SF, Coleman JS, Arnone JA (2003) *In situ* litter decomposition and litter quality in a Mojave Desert ecosystem: effects of elevated atmospheric CO<sub>2</sub> and interannual climate variability. *Global Change Biology*, **9**, 1223–1233.
- Wedin DA, Tilman D (1996) Influence of nitrogen loading and species composition on the carbon balance of grasslands. *Science*, **274**, 1720–1723.
- Zak DR, Holmes W, Finzi AC, Norby RJ, Schlesinger WH (2003) Soil nitrogen cycling under elevated CO<sub>2</sub>: a synthesis of forest face experiments. *Ecological Applications*, **13**, 1508–1514.