

## Effects of fire frequency on plant species in oak savanna in east-central Minnesota<sup>1</sup>

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TESTER, J. R. (Department of Ecology, Evolution and Behavior, University of Minnesota, St. Paul, MN 55108). Effects of fire frequency on plant species in oak savanna in east-central Minnesota. Bull. Torrey Bot. Club 123: 304–308, 1996.—From 1964 through 1984, nine forest sites, ranging in size from 2.6 to 27.5 ha, were each burned from two to 19 times. Percent cover of 13 of 14 true-prairie grasses was positively correlated with burn frequency. Of these, eight have C4 and six have C3 photosynthetic pathways. Cover of 34 of 39 true-prairie forbs, and of *Amorpha canescens*, increased with frequency of burning. All except *Aster oolentangiensis* are C3 species. Cover of six of seven native, not prairie, species, all C3, decreased with increasing burn frequency. These data suggest that the adaptation of true-prairie species to repeated burning outweighs the effects of their photosynthetic pathways.

Key words: oak savanna, prescribed burning, prairie plants.

The role of fire in maintenance of tallgrass prairie and savanna ecosystems is well documented (reviewed in Collins and Wallace 1990; Hulbert 1986; Johnson 1993). In general, fire suppresses shrubs and trees and favors native prairie species. Responses of plant species to burning, however, are related to season of burning (Howe 1994a, 1994b), intensity and frequency of fires (Gibson 1988; Tester 1989; White 1986). However, plant species respond in an individualistic manner. Thus, use of prescribed burning by resource managers must be based on knowledge of how individual species will respond to the treatment. This paper reports on the response of certain plant species over a 20-year period to a program of prescribed fires that was implemented in 1964 at the Cedar Creek Natural History Area to restore oak savanna from oak forest.

**Methods.** In 1964, nine forest sites, ranging in size from 2.6 to 27.5 ha, were established and randomly assigned for the application of different frequencies of prescribed burns (Irving 1970). The number of burns at a given site for the 20-year period ranged from two to 19. Dates, characteristics of burns, and design limitations are given in Tester (1989). Unfortunately, no data were collected on these nine sites prior to initiation of the burning regimen in 1964.

These sites plus three unburned sites in adja-

cent oak forest believed to be typical of the entire study area were used for the present study. All 12 sites are within a 320-ha block and have similar topography. In 1984, percent cover of litter, bare ground, and each shrub and herb species was visually estimated in 24 1.0-m × 0.5-m quadrats, spaced uniformly in the sample area, following Inouye et al. (1987). Each species was placed in one of three categories; (1) typically occurring in true (tallgrass) prairie, (2) native to North America but not typically found in prairie, or (3) introduced from another continent, using data from Gleason and Cronquist (1963), Curtis (1959), and personal communications with E. Cushing, B. Delaney, and T. Morley. Species also were categorized by photosynthetic pathways following Waller and Lewis (1979) and E. Wardenaar (pers. comm.). Nomenclature follows Ownbey and Morley (1991). Regression analyses were performed with number of burns as the independent variable and average of percent cover as the dependent variable. No a priori criteria were available to identify outliers; therefore, all data points were included for all species.

**Results and Discussion.** Litter comprised from 28.9 to 85.0 percent of the cover in the 12 plots and bare ground varied from <1.0 to 38.5 percent. A total of 104 shrub and herb species were identified on the study area. Cover estimates of 13 of 14 true-prairie grasses were positively correlated with burn frequency (Table 1), with the highest significance levels noted for *Panicum praecociosus*, *Sorghastrum nutans* (Fig. 1), *Stipa spartea*, *Bouteloua hirsuta*, *Schizachyrium scoparius*, *Panicum virgatum*, and *An-*

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1. The first part of the document discusses the importance of maintaining accurate records of all transactions and activities. It emphasizes that this is crucial for ensuring transparency and accountability in the organization's operations.

2. The second part of the document outlines the various methods and tools used to collect and analyze data. It highlights the need for consistent and reliable data collection processes to support informed decision-making.

3. The third part of the document focuses on the role of technology in data management and analysis. It discusses how modern software solutions can streamline data collection, storage, and reporting, thereby improving efficiency and accuracy.

4. The fourth part of the document addresses the challenges associated with data management, such as data quality, security, and privacy. It provides strategies to mitigate these risks and ensure that data is used responsibly and ethically.

5. The fifth part of the document concludes by summarizing the key findings and recommendations. It stresses the importance of ongoing monitoring and evaluation to ensure that data management practices remain effective and aligned with the organization's goals.

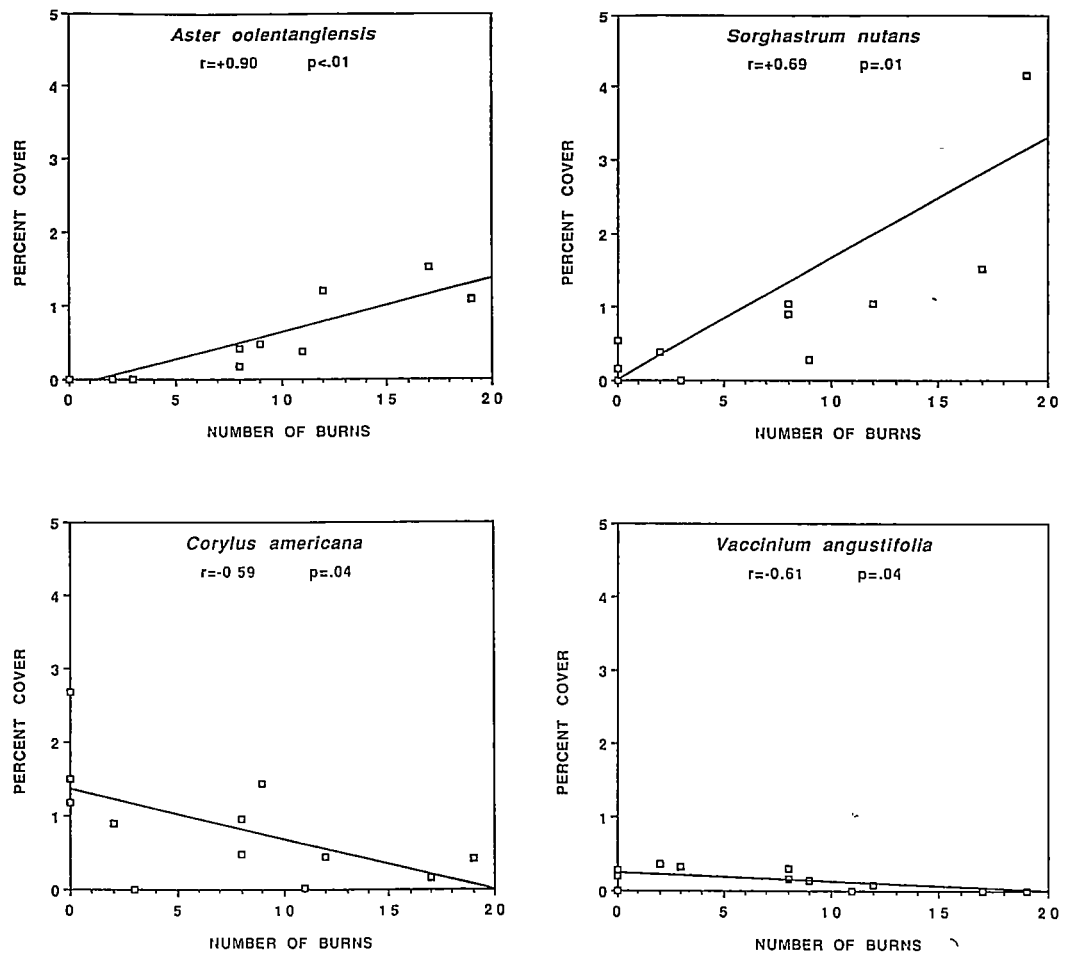


Fig. 1. Linear regressions showing effect of number of burns on percent cover of *Aster oolentangiensis*, *Sorghastrum nutans*, *Corylus americana*, and *Vaccinium angustifolia*.

*dropogon gerardi*. Six have C3 and eight have C4 photosynthetic pathways.

Increase in C4 grasses has been reported from numerous studies (Gibson 1989; Hadley and Kieckhefer 1963; Svedarsky et al. 1986; and others). However, increased cover of C3 grasses is somewhat surprising. Anderson et al. (1970), Gibson and Hulbert (1987), and Glenn-Lewin et al. (1990) stated that C3 grasses generally decreased after spring fire, and Howe (1994a, 1994b) emphasized that spring burns are likely to suppress C3 and favor C4 species. Anderson and Bailey (1980) reported that annual burning for 24 years reduced cover of *Stipa spartea*, a C3 species, in Alberta.

Cover of two introduced grasses, *Poa pratensis* (C3) and *Setaria lutescens* (C4), showed increases in cover with burn frequency and two

other introduced species, *Agropyron repens* (C3) and *Bromus inermis* (C3), showed decreases. However, none of the *P*-values were less than 0.23.

Linear regressions showing the effect of number of fires on percent cover of *Corylus americana* and *Vaccinium angustifolia* are shown in Fig. 1. Cover estimates of six of the seven native, non-prairie species with  $P < 0.10$ , including the above, were negatively correlated with burn frequency. This result is not surprising since these species are typical of Minnesota forests where fire would have been less frequent (Tester 1995). I cannot explain the one exception, *Parthenocissus vitacea*, which significantly increased with burn frequency. All native, non-prairie species in Table 1 have C3 photosynthetic pathways.

Table 1. Linear regression showing effects of burn frequency on percent cover of shrubs and herbs. Ecological origin, photosynthetic pathway, and life form are given for each species.

	Origin <sup>a</sup>	Photo-synthetic pathway	Life form <sup>b</sup>	r	P	P < 0.10
<i>Achillea millefolium</i>	T	C3	F	0.028	0.931	
<i>Agropyron repens</i>	I	C3	G	-0.372	0.233	
<i>Ambrosia coronopifolia</i>	T	C3	F	0.334	0.288	
<i>Amorpha canescens</i>	T	C3	SH	0.572	0.052	†
<i>Amphicarpa bracteata</i>	N	C3	F	0.141	0.663	
<i>Andropogon gerardi</i>	T	C4	G	0.578	0.049	†
<i>Anemone cylindrica</i>	T	C3	F	0.179	0.577	
<i>Antennaria plantagenifolia</i>	T	C3	F	0.281	0.376	
<i>Apocynum cannabinum</i>	N	C3	SH	-0.355	0.258	
<i>Arenaria lateriflora</i>	N	C3	F	-0.377	0.227	
<i>Artemisia caudata</i>	T	C3	F	0.171	0.594	
<i>Artemisia ludoviciana</i>	T	C3	F	0.464	0.129	
<i>Asclepias syriaca</i>	N	C3	F	-0.208	0.516	
<i>Asclepias tuberosa</i>	T	C3	F	0.201	0.531	
<i>Aster oolentangiensis</i>	T	C4	F	0.898	0.000	†
<i>Bouteloua hirsuta</i>	T	C4	G	0.596	0.041	*
<i>Bromus inermis</i>	I	C3	G	-0.002	0.995	
<i>Calamagrostis canadensis</i>	T	C3	G	0.223	0.486	
<i>Calamovilfa longifolia</i>	T	C4	G	0.478	0.116	
<i>Campanula rotundifolia</i>	T	C3	F	0.219	0.494	
<i>Ceanothus americanus</i>	T	C3	SH	0.028	0.931	
<i>Celastrus scandens</i>	N	C3	SH	0.209	0.514	
<i>Chenopodium alba</i>	I	C3	F	-0.351	0.264	
<i>Commandra richardsiana</i>	T	C3	F	0.383	0.219	
<i>Coreopsis palmata</i>	N	C3	F	0.460	0.133	
<i>Cornus racemosa</i>	N	C3	SH	0.028	0.931	
<i>Cornus stolonifera</i>	N	C3	SH	-0.233	0.467	
<i>Corylus americana</i>	N	C3	SH	-0.588	0.044	*
<i>Crepis tectorum</i>	I	?	F	0.342	0.277	
<i>Desmodium canadense</i>	N	C3	F	-0.005	0.989	
<i>Elymus canadensis</i>	T	C3	G	0.028	0.931	
<i>Equisetum laevigatum</i>	T	C3	N	-0.285	0.369	
<i>Eragrostis spectabilis</i>	T	C4	G	-0.038	0.908	
<i>Erigeron canadensis</i>	N	C3	F	0.307	0.332	
<i>Erigeron strigosus</i>	N	C3	F	0.458	0.134	
<i>Euphorbia corollata</i>	T	C3	F	0.458	0.134	
<i>Fragaria virginiana</i>	T	C3	F	-0.132	0.682	
<i>Galium aparine</i>	T	C3	F	0.076	0.815	
<i>Galium boreale</i>	N	C3	F	0.121	0.707	
<i>Galium triflorum</i>	N	C3	F	-0.515	0.086	*
<i>Helianthemum bicknellii</i>	T	C3	F	0.694	0.012	*
<i>Helianthus laetiflorus</i>	T	C3	F	0.567	0.055	†
<i>Juniperus communis</i>	T	C3	SH	-0.470	0.124	
<i>Lactuca canadensis</i>	T	C3	F	0.009	0.977	
<i>Lathyrus venosus</i>	N	C3	F	0.075	0.818	
<i>Lechea stricta</i>	T	C3	F	-0.089	0.782	
<i>Lespedeza capitata</i>	T	C3	F	0.224	0.485	
<i>Lithospermum canescens</i>	T	C3	F	0.615	0.033	†
<i>Lithospermum carolinense</i>	T	C3	F	-0.005	0.989	
<i>Lychnis alba</i>	I	C3	F	0.268	0.399	
<i>Lysimachia ciliata</i>	N	C3	F	0.028	0.931	
<i>Maianthemum canadense</i>	N	C3	F	-0.220	0.493	
<i>Monarda fistulosa</i>	N	C3	F	-0.292	0.357	
<i>Muhlenbergia racemosa</i>	T	C4	G	0.038	0.907	
<i>Oenothera biennis</i>	T	C3	F	0.355	0.258	
<i>Oxybaphus hirsutus</i>	T	C3	F	0.193	0.548	
<i>Panicum oligosanthes</i>	T	C3	G	0.524	0.080	†
<i>Panicum perlongum</i>	T	C3	G	0.548	0.065	**
<i>Panicum praeccocius</i>	T	C3	G	0.765	0.004	**
<i>Panicum virgatum</i>	T	C4	G	0.585	0.046	†
<i>Parthenocissus inserta</i>	N	C3	SH	0.391	0.209	
<i>Parthenocissus vitacea</i>	N	C3	SH	0.678	0.015	†

Table 1. Continued

	Origin <sup>a</sup>	Photo-synthetic pathway	Life form <sup>b</sup>	r	P	P < 0.10
<i>Pedicularis canadensis</i>	T	C3	F	0.219	0.494	
<i>Physalis virginiana</i>	T	C3	F	0.406	0.190	
<i>Poa pratensis</i>	I	C3	G	0.169	0.600	
<i>Polygala polygama</i>	T	C3	F	0.587	0.045	*
<i>Polygonatum biflorum</i>	N	C3	F	0.079	0.807	
<i>Polygonum convolvulus</i>	I	C3	F	0.029	0.930	
<i>Potentilla simplex</i>	N	C3	F	0.083	0.799	
<i>Prunus americana</i>	N	C3	SH	-0.355	0.258	
<i>Prunus virginiana</i>	N	C3	SH	-0.512	0.089	*
<i>Ranunculus rhomboideus</i>	T	C3	F	0.137	0.671	
<i>Rhus glabra</i>	T	C3	SH	0.130	0.688	
<i>Rhus radicans</i>	T	C3	SH	0.390	0.210	
<i>Rosa arkansana</i>	T	C3	SH	0.232	0.468	
<i>Rubus sp</i>	N	C3	SH	0.068	0.835	
<i>Rudbeckia serotina</i>	T	C3	F	0.045	0.890	
<i>Rumex acetosella</i>	I	C3	F	-0.131	0.685	
<i>Salix sp</i>	T	C3	SH	-0.179	0.578	
<i>Schizachyrium scoparium</i>	T	C4	G	0.590	0.044	*
<i>Scutellaria parvula</i>	T	C3	F	0.687	0.014	†
<i>Senecio pauperculus</i>	T	C3	F	0.488	0.108	
<i>Setaria lutescens</i>	I	C4	G	0.246	0.441	
<i>Sisyrinchium campestre</i>	T	C3	F	0.554	0.062	*
<i>Smilacina racemosa</i>	N	C3	F	0.007	0.984	
<i>Smilacina stellata</i>	T	C3	F	0.188	0.559	
<i>Smilax herbacea</i>	N	C3	F	0.259	0.416	
<i>Solanum dalcamara</i>	I	C3	F	-0.211	0.510	
<i>Solidago gigantea</i>	T	C3	F	-0.016	0.959	
<i>Solidago graminifolia</i>	T	C3	F	0.455	0.137	
<i>Solidago missouriensis</i>	T	C3	F	0.438	0.155	
<i>Solidago nemoralis</i>	T	C3	F	0.560	0.058	*
<i>Solidago rigida</i>	T	C3	F	0.142	0.661	
<i>Sorghastrum nutans</i>	T	C4	G	0.694	0.012	*
<i>Stachys palustris</i>	T	C3	F	0.173	0.590	
<i>Stellaria longifolia</i>	N	C3	F	-0.355	0.258	
<i>Stipa spartea</i>	T	C3	G	0.834	0.001	*
<i>Trientalis borealis</i>	N	C3	F	-0.529	0.077	*
<i>Trifolium sp</i>	I	C3	F	-0.211	0.510	
<i>Uvularia sessifolia</i>	N	C3	F	-0.355	0.258	
<i>Vaccinium angustifolia</i>	N	C3	SH	-0.605	0.037	*
<i>Viola pedatifida</i>	T	C3	F	0.033	0.918	
<i>Viola sagittata</i>	T	C3	F	-0.350	0.264	
<i>Vitis riparia</i>	N	C3	SH	-0.508	0.092	†

<sup>a</sup> T—true prairie, N—native, non-prairie, I—introduced.

<sup>b</sup> F—forb, G—grass, N—non-vascular, SH—shrub.

All true-prairie forbs with  $P < 0.10$ , 34 of the 39 true-prairie forbs listed in Table 1, and *Amarpha canescens*, a true-prairie shrub, increased with burn frequency. All except *Aster oolentangiensis* (Fig. 1) are C3. Increase in *Amarpha canescens* was reported by Abrams and Hulbert (1987), but Gibson et al. (1993) found that cover of *Amarpha canescens* was reduced by burning. Gibson et al. (1993) reported that cover of forbs was reduced by burning and Gibson and Hulbert (1987) indicated that C3 species were reduced and C4 species favored by spring burning. Published information on the response of most forbs

listed in Table 1 to fire frequency is lacking. However, Kline (1986) found that *Solidago nemoralis* decreased under annual burning.

Management of tallgrass prairie is dependent on knowledge of how individual species will respond to treatments such as prescribed burning. Data presented here on the response of selected native true-prairie and native, non-prairie species to fire frequency indicate that the true prairie species are well adapted to fire, and that this adaptation appears to outweigh the effects of the photosynthetic pathway on response of the individual species.

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